

# A LOW-COST AND EASY TO USE SORTING DEVICE FOR THE SEPARATION OF EXTRACELLULAR VESICLES FROM COMPLEX FLUIDS

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# A LOW-COST AND EASY TO USE SORTING DEVICE FOR THE SEPARATION OF EXTRACELLULAR VESICLES FROM COMPLEX FLUIDS













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#### CONTEXT

Identifying and characterizing Extracellular vesicles (EVs) in biofluids could lead to unprecedented diagnosis/prognosis assays for a wide set of pathologies.

Owe to their complexity in size, origin, membrane markers, there is currently no ideal technology available yet to sort, to quantify and characterize cell-derived microvesicles, and fully relate their structure and functions which have high potential in clinical diagnostics and early treatment of life threatening diseases such as cancer, cardio vascular diseases....

#### SEPARATION PRINCIPLE: HYDRODYNAMIC FILTRATION

Exclusion of large particle from the streamlines located close to a wall

At the channel crossing, a particle flowing next to the sidewall: Sample ==>

- radius < R<sub>c</sub> → flow in **lateral channel**
- radius > R<sub>c</sub> → stay in the main channel

#### Advantages and drawbacks:

- ✓ Passive method ⇔ compatible with various applications
- ✓ Robustness ⇔ less prone to clogging than filter-based geometries
- ✓ Reliability ⇔ independent of flow and pressure drop

### LAB ON CHIP: DESIGN AND MANUFACTURING

Choosing the appropriate dimensions of the channels, we designed and fabricated chips enabling the separation of vesicles at tunable cutoff (150 - 900 nm).

w	h	L	Flow rate
(µm)	(µm)	(cm)	(µL/s)
10	10	0.12 to 7.59	0.035

111111

"Filtered"

EVs

Lateral

channel

"No filtered"

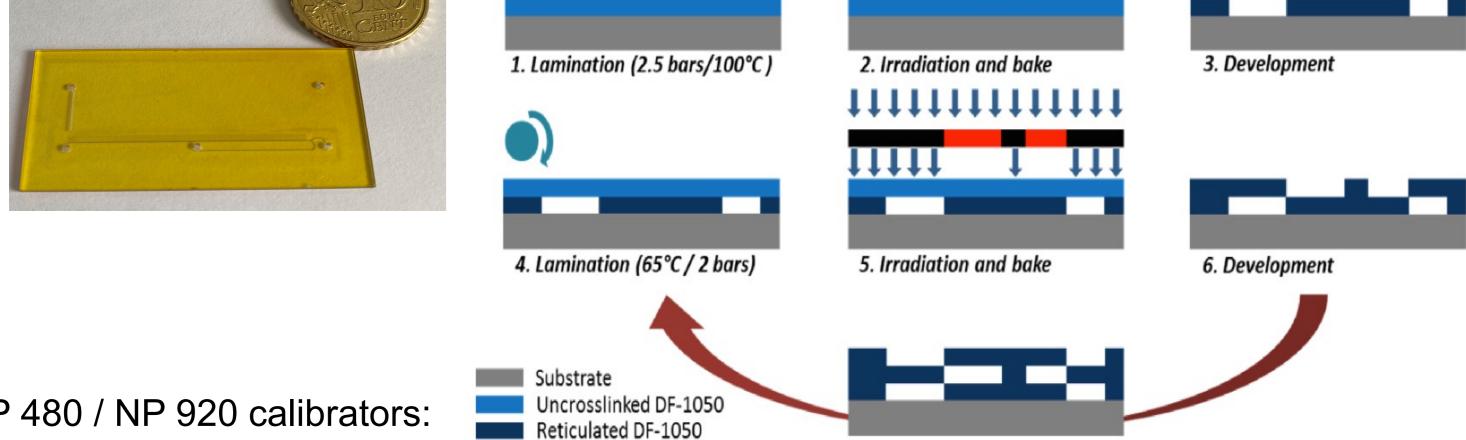
EVs

Main channel

#### Fabrication process: successive lamination and photolithography

Dry Film: manufacturable process by laminating dry photosensible films

- Low cost
- Compatible with micronanotechnologies
- Biocompatible



### Samples preparations

- Development of NP 140 / NP 480 / NP 920 calibrators: Biofunctionalization by proteins(OVA, CAS)
- Approx. 3500 CAS / NP 140 and 17000 OVA / NP 480
- Selective trapping verification by anti-CAS and anti-OVA

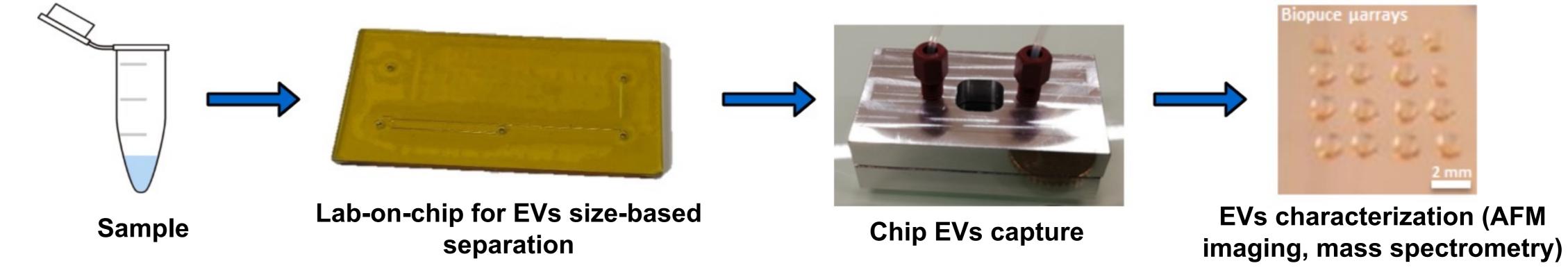
NPs	Diametre	Proteins	Immunocapture
NP 140	140 nm	Casein	anti-CAS
NP 480	480 nm	Ovalbumin	anti-OVA
NP 920	920 nm	_	_

Synthetic solution	NPs in PBS-Tween 20 (0.1%)	[NPs] = 1.10 <sup>8</sup> part/mL	
Blood sample	Plasma diluted x 10	[NPs] in PBS- Tween20: 2.10 <sup>6</sup> to 1.10 <sup>8</sup> part/mL	
	Blood diluted X 10 and x 100		

## **APPROACH**

We propose to couple a microfluidic device (LOC) enabling to separate particles from a complex sample to a device allowing the capture of particles on specific spots and their analysis.

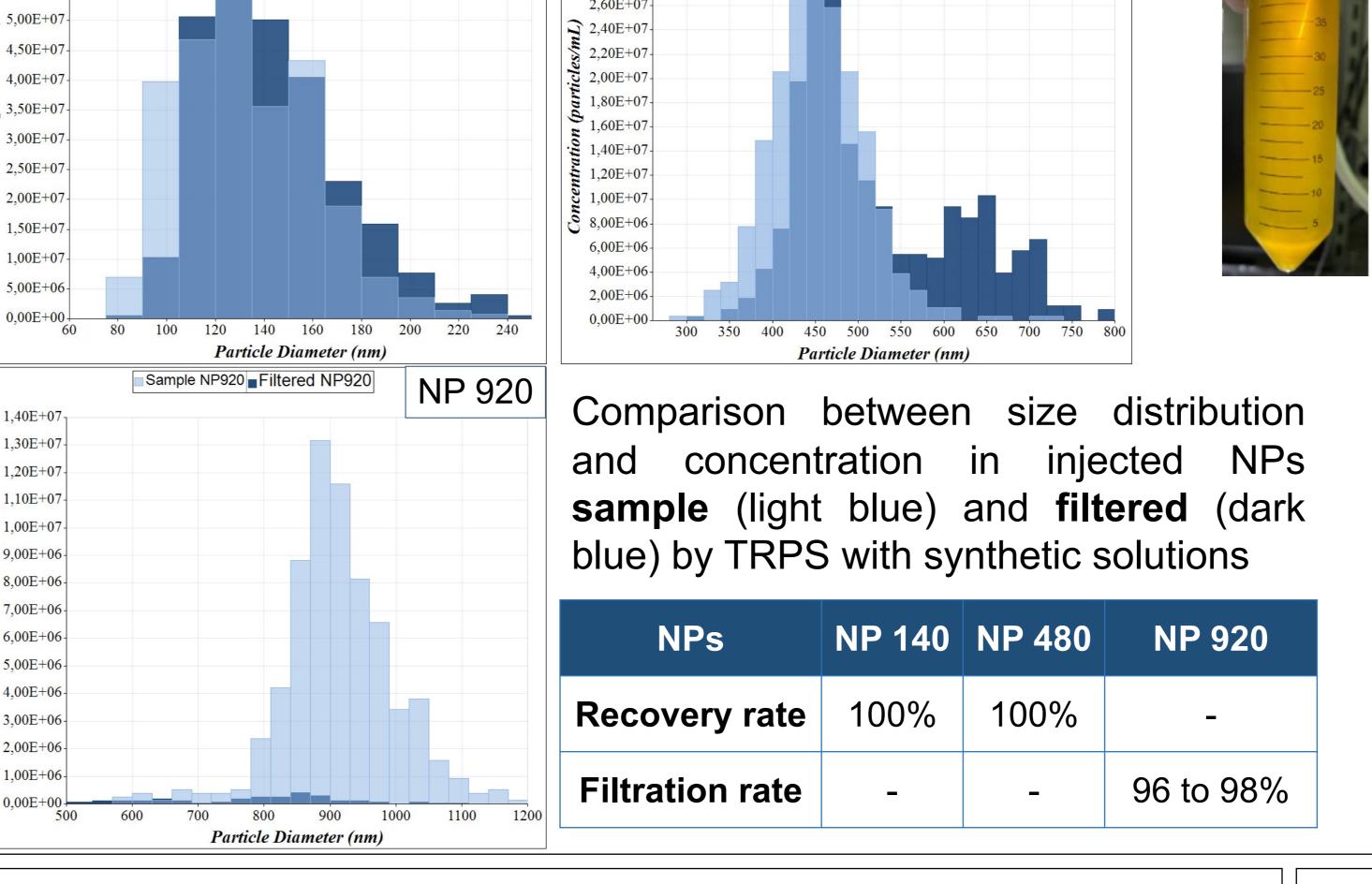
These methods allow obtaining important information on a chip about nanoscale vesicles in terms of size, membrane markers expression, and structure-function properties.

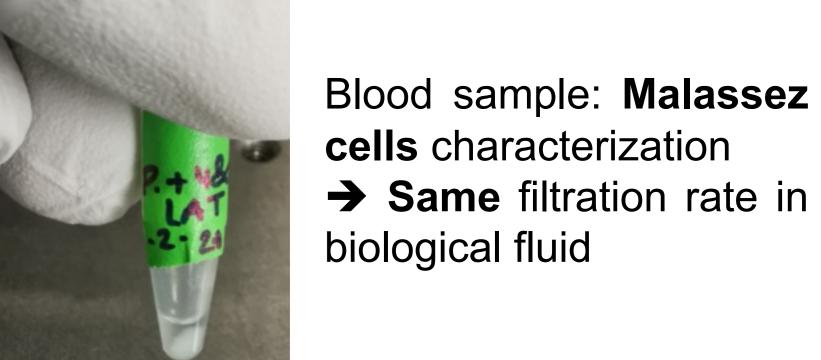


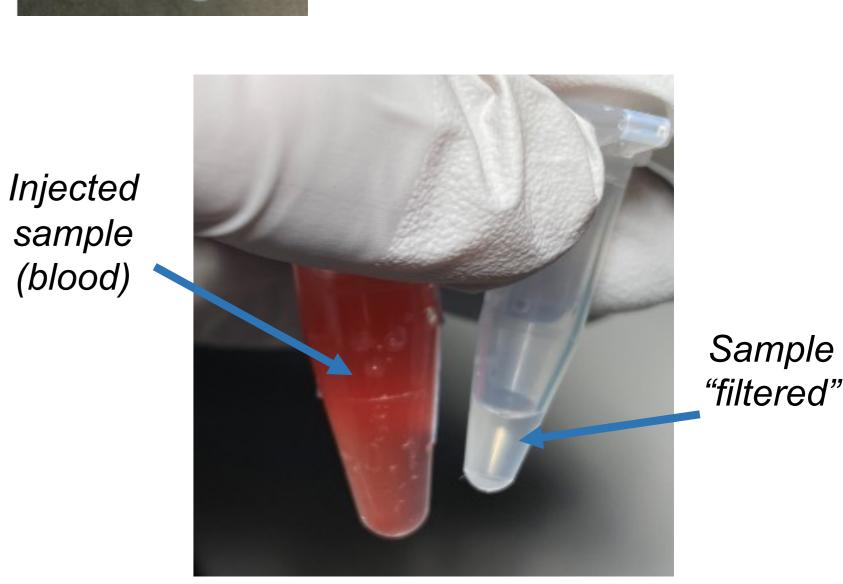
NP 480

### CHARACTERIZATION AND VALIDATION OF LAB ON CHIP

NP 140







## CONCLUSION

- Proof-on-concept: NP 140 and 480 were filtered while NP 920 were find in the unfiltered sample.
- Recovery conditions: 35 µL sample filtered in 30 minutes.
- With NPs in PBS-Tween20 and NPs in blood sample, we obtained the same recovery rate (100%) and filtration rate (96 to 98%).
- We showed high efficiency separation with NPs and biological fluids did **not affect** the separation.
- We are also improving the performances of the devices (filtration flow rate, filtered volume) and are coupling it to an immuno-trapping device in order to provide an integrated EVs sorting and capture bio-analytical system.

#### REFERENCES

(blood)

Yamada, M., & Seki, M. (2005). Hydrodynamic filtration for on-chip particle concentration and classification utilizing microfluidics. Lab on a Chip, 5(11), 1233-1239. Fouet, M., et al. "Filter-less submicron hydrodynamic size sorting." Lab on a Chip 16,4 (2016): 720-733. Courson, R, et al. "Low-cost multilevel microchannel lab on chip: DF-1000 series dry film photoresist as a promising enabler." RSC Advances 4,97 (2014): 54847-54853.

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